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# ORK1, a potassium-selective leak channel with two pore domains cloned from *Drosophila melanogaster* by expression in *Saccharomyces cerevisiae*

(K<sup>+</sup> channel/two P domains/yeast/muscles/nerves)

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**ABSTRACT** A K<sup>+</sup> channel gene has been cloned from *Drosophila melanogaster* by complementation in *Saccharomyces cerevisiae* cells defective for K<sup>+</sup> uptake. Naturally expressed in the neuromuscular tissues of adult flies, this gene confers K<sup>+</sup> transport capacity on yeast cells when heterologously expressed. In *Xenopus laevis* oocytes, expression yields an ungated K<sup>+</sup>-selective current whose attributes resemble the “leak” conductance thought to mediate the resting potential of vertebrate myelinated neurons but whose molecular nature has long remained elusive. The predicted protein has two pore (P) domains and four membrane-spanning helices and is a member of a newly recognized K<sup>+</sup> channel family. Expression of the channel in flies and yeast cells makes feasible studies of structure and *in vivo* function using genetic approaches that are not possible in higher animals.

Potassium channels comprise a large group of integral membrane proteins that determine the electrical properties of neurons and muscle fibers (1). They assume this central role in cellular physiology through regulation of action potential duration, firing patterns, and resting membrane potential. Distinguished individually by how they open and close and the magnitude of their single channel conductances, K<sup>+</sup> channel subunits are unified by a shared pore-forming motif, the P domain (2, 3). Molecular cloning has revealed genes in *Drosophila melanogaster* and mammals encoding a superfamily of closely related K<sup>+</sup> channels whose subunits contain a single P domain and six or more membrane-spanning segments (4–10). Another K<sup>+</sup> channel superfamily, the inward rectifiers, is characterized by a single P domain and just two transmembrane segments (11, 12). The first K<sup>+</sup> channel possessing two P domains in one contiguous polypeptide, Tok1, was cloned from the budding yeast *Saccharomyces cerevisiae* and has eight predicted membrane-spanning  $\alpha$ -helices (13). More recently, a second family of K<sup>+</sup> channels with two P domains and just four predicted transmembrane segments was expressed (14), confirming predictions based on data base analysis (13, 15, 16).

Here, we describe the expression cloning from *D. melanogaster* of an example of this later type of two P domain K<sup>+</sup> channel with four predicted transmembrane segments. The gene for the channel is located on chromosome 1 and gains wide expression in the excitable tissues of adult flies. Heterologous expression of the gene in yeast cells defective in K<sup>+</sup> transport restores K<sup>+</sup> uptake ability and allows cells to grow in medium containing micromolar concentrations of potassium. Gene expression in *Xenopus laevis* oocytes bathed in physiological solutions yields K<sup>+</sup>-selective currents that are instantaneous with changes in voltage and are outwardly rectifying. However, increased external K<sup>+</sup> concentration ([K]<sub>o</sub>) significantly alters this current–voltage relationship; as [K]<sub>o</sub> rises, rectification is lost and K<sup>+</sup> ions flow inward at

hyperpolarized potentials. This sensitivity to [K]<sub>o</sub> appears intrinsic to the channel, since it approximates expectations from constant field theory for simple electrodiffusion through an open K<sup>+</sup>-selective pore (17, 18). That the channel directly underlies all these events is supported by the observation that K<sup>+</sup> uptake, yeast cell growth, and oocyte currents are similarly sensitive to barium and resistant to tetraethylammonium (TEA).

While “leak conductances” have long been thought to be basic to neural function (19, 20), efforts to characterize them in dissected nerves have yielded inconsistent results (1). In function, the channel we describe here resembles the rectifying “leak” conductance postulated to underlie the resting potential of myelinated nerves (21) and is, to our knowledge, the first cloned channel to exhibit such behavior. Study of the channel’s influence on fly physiology in wild-type and mutant form may serve to clarify the role of leak conductances in neural function. Investigation of the channel in yeast cells allows application of genetic methods to assess the molecular basis for its function as well as high-throughput screens for isolation of channel modulators. The new gene and its channel product have been named ORK1, as an abbreviation for open rectifier K<sup>+</sup> channel.

## MATERIALS AND METHODS

**Molecular Biology.** Cloning of ORK1 by complementation. Plasmid DNA was isolated from a  $\lambda$ YES cDNA expression library made from third-instar larval mRNA (22). Library plasmids were introduced into *S. cerevisiae* strain CY162 by using the lithium acetate method, and 10<sup>5</sup> transformants were screened for complementation on low K<sup>+</sup> as described (23). Plasmids isolated from colonies viable on low K<sup>+</sup> were used to transform CY162 to confirm their complementing properties. One positive plasmid contained a complementing 2.4-kb *Xho*I fragment which was transferred to the *Xho*I sites of pYES2 (Invitrogen), forming pORK1, and to a variant of pRS426 (Stratagene) bearing the ADHI promoter (24), forming pRAD-ORK1. Yeast media and methods were conventional; SC, complete synthetic medium, uracil-deficient with 3% galactose and  $\leq 1$  mM KCl; RPD, arginine phosphate dextrose medium, uracil-deficient and  $\leq 1$  mM KCl (25). *S. cerevisiae* strain CY162 (*ura3 trk1 trk2*) and the plasmid pKAT1 were obtained from Rick Gaber and have been described (23).

**DNA sequence analysis.** Automated DNA sequence analysis was performed with Prism DNA sequencing kits (Applied Biosystems). Sequences were analyzed by using GeneWorks software (Intelligenetics). Quality scores were generated by the Gap Program of the Wisconsin Package (Version 8), Genetics Computer Group (26). A combined transcription–

Abbreviations: TEA, tetraethylammonium; ORF, open reading frame. Data deposition: The sequence reported in this paper has been deposited in the GenBank data base (accession no. U55321).

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translation protocol (Promega) with [<sup>35</sup>S]methionine (NEN) was used. SDS/polyacrylamide gels were impregnated with soluble fluor (Amplify; NEN), and exposed to x-ray film (XAR-5, Kodak) after drying.

**Rubidium uptake assay.** CY162 bearing pRAD-ORK1 or pRAD were grown overnight in RPD medium containing 100 mM KCl and no uracil, washed in distilled water, incubated in 10 mM calcium Hepes buffer (pH 6.8) for 1 hr, washed, and resuspended in buffer at  $2 \times 10^9$  cells per ml. A 50- $\mu$ l aliquot of the yeast suspension was mixed with 50  $\mu$ l of buffer containing 5 kBq of <sup>86</sup>RbCl (Amersham) and the indicated nonradioactive chloride salt. After 45 min, cells were concentrated on a 0.45- $\mu$ m-pore HVLP Millipore filter and washed, and radioactivity was measured in a liquid scintillation counter. Rubidium uptake rates remained linear for over 60 min in calcium Hepes buffer containing 50  $\mu$ M RbCl (not shown). Uptake by cells expressing pYES2-KAT1 was roughly 2-fold more efficient than in those expressing pRAD-ORK1 in buffer containing 50  $\mu$ M RbCl (not shown).

**Synthesis of ORK1 cRNA for microinjection.** Synthetic DNA duplexes were used to ligate the DNA sequence encoding ORK1 into the *Nco*I and *Bsp*el sites of pSD (27), creating pSD-ORK1. Complementary RNA synthesis was catalyzed by T7 RNA polymerase *in vitro* using pSD-ORK1 as template after linearization with *Not*I. Transcript concentration was estimated spectrophotometrically and aliquots were stored at  $-80^\circ\text{C}$ .

**Electrophysiology.** Expression of ORK1 in *X. laevis* oocytes. Oocytes were isolated from *X. laevis* (Nasco, Atkinson, WI), defolliculated by collagenase treatment, and injected the next day with 46 nl of cRNA solution containing 1–5 ng of transcript. Whole cell currents were measured 1–3 days after cRNA injection by two-electrode voltage clamp using an Oocyte Clamp (Warner). Data were filtered at 1 kHz and sampled at 4 kHz. Electrodes contained 3 M KCl and had resistances of 0.3–1 M $\Omega$ . Recordings were performed under constant perfusion at room temperature. Bath solutions were 5 mM KCl solution (in mM: 95 NaCl, 5 KCl, 1 MgCl<sub>2</sub>, 0.3 CaCl<sub>2</sub>, 5 Hepes, pH 7.6); or 10, 20, 50, and 100 mM KCl solutions in which KCl is isotopically substituted for NaCl. For aspartate substitution, KCl and NaCl were replaced with potassium aspartate and sodium aspartate, respectively; for NMG substitution, NaCl was replaced with *N*-methylglucamine chloride.

## RESULTS

**Molecular Cloning of ORK1 by Functional Expression in *S. cerevisiae*.** In an effort to clone novel K<sup>+</sup> channels, we employed a yeast complementation method used previously to isolate channels and transporters from plants (23, 28, 29). A yeast mutant, CY162, lacking high-affinity K<sup>+</sup> uptake (23) was transformed with a *D. melanogaster* cDNA expression library made from third-instar larval mRNA (22). A single plasmid clone conferred upon these cells the ability to grow on low-K<sup>+</sup> medium. The growth of yeast cells bearing this plasmid was comparable to that of control strains containing the inwardly-rectifying K<sup>+</sup> channel of the plant *Arabidopsis thaliana*, KAT1 (23), and was much greater than that of CY162 cells containing vector alone (Fig. 1). Subcloning identified a 2422-bp *Xho*I fragment that conferred growth on low-K<sup>+</sup> medium.

DNA sequencing of the 2422-bp fragment revealed a single long open reading frame (ORF) encoding a predicted protein of 618 amino acids (Fig. 2a). The start codon appears to be the first in-frame ATG, within the strong translation initiation sequence AGCATGT (31, 32). Within the proposed 3' untranslated region is a consensus sequence for polyadenylation, AATCAA (33), 122 bases downstream of the translational stop sequence. Hydrophobicity analysis of the predicted protein shows four stretches that could span the membrane in  $\alpha$ -helical conformation (M1–M4) (Fig. 2b). Two motifs bearing close resemblance to the P domains of known K<sup>+</sup> channels are in the ORF (Fig. 2c); the first (P1) is flanked by hydrophobic segments M1 and M2, the second (P2), by M3 and M4. The predicted mass of the protein ( $\approx$ 68

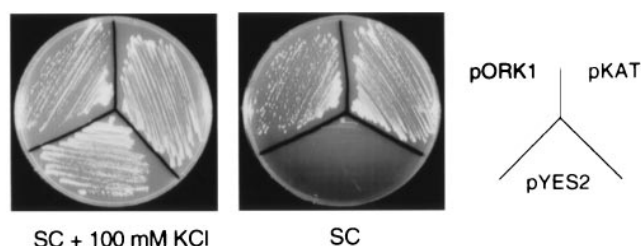


FIG. 1. Molecular cloning of ORK1 by complementation. ORK1 expression allows growth of yeast cells deficient in K<sup>+</sup> transport; CY162 yeast cells were transformed with the designated plasmids, streaked on plates of uracil-deficient synthetic complete medium (SC), or SC containing 100 mM KCl and incubated for 40 hr at 30°C.

kDa) and a consensus site for N-glycosylation are consistent with products observed upon translation of ORK1 cRNA *in vitro* with and without microsomes (Fig. 2d). In the absence of a signal sequence, these data suggest a membrane topology in which the N and C termini are inside the cell and residue 58 (between M1 and M2) is glycosylated and thus external (Fig. 2e).

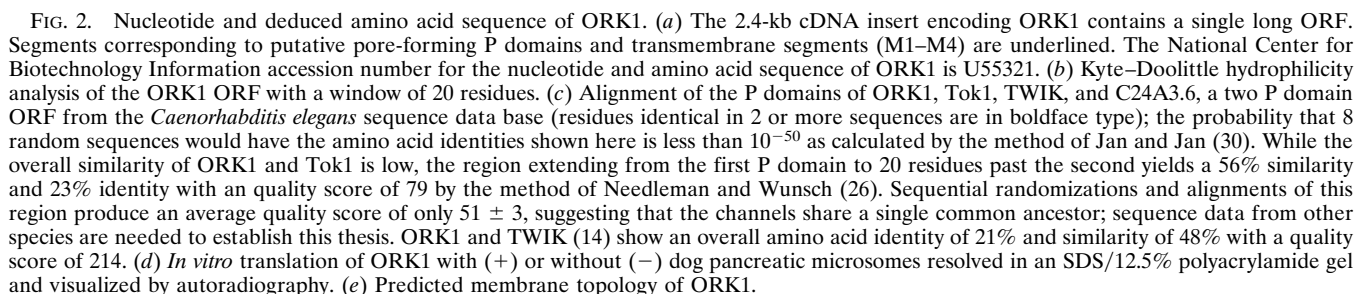
Northern blot analysis using the 2422-bp *Xho*I fragment as the probe reveals a 2.8-kb species of poly(A)<sup>+</sup> mRNA from *D. melanogaster* adults (Fig. 3a) that is also present at low levels in larvae and embryos (not shown). *In situ* hybridization to polytene chromosomal preparations localizes ORK1 to band 1-10A1-2 (Fig. 3b); several lethal mutations map to this band (34). *In situ* RNA-RNA hybridization using a 750-bp antisense fragment reveals that expression of ORK1 mRNA is widespread, with particular prominence in muscle, brain, and ovary of adult flies (Fig. 3c).

**ORK1 Mediates Barium-Sensitive Uptake of K<sup>+</sup> When Expressed in Yeast Cells.** CY162 yeast cells grow poorly on low-K<sup>+</sup> medium unless they express an exogenous protein that mediates K<sup>+</sup> transport, such as the plant K<sup>+</sup> channel KAT1 (Fig. 1) (23). Consistent with the idea that ORK1 is also an ion channel, CY162 cells constitutively expressing ORK1 grow on low-K<sup>+</sup> medium (RPD containing 1 mM KCl) and their growth is inhibited by the common K<sup>+</sup> channel blocker barium (Fig. 4a). Cell growth resulting from ORK1 expression differs in its pharmacology from that based on KAT1—only cells employing the latter are markedly sensitive to TEA and cesium (Fig. 4a). This difference suggests that the two proteins facilitate transport directly rather than by activating an endogenous transport protein.

To directly assess ORK1-mediated ion flux into yeast cells, uptake of the K<sup>+</sup> congener <sup>86</sup>Rb<sup>+</sup> was evaluated. Constitutive expression of ORK1 in CY162 cells results in <sup>86</sup>Rb<sup>+</sup> uptake rates comparable to those observed with KAT1 and much higher than those seen with vector alone (Fig. 4b). ORK1-mediated uptake of <sup>86</sup>Rb<sup>+</sup> is concentration dependent and, like cell growth, is sensitive to barium but not TEA (Fig. 4c).

**ORK1 Mediates K<sup>+</sup>-Selective Currents When Expressed in *X. laevis* Oocytes.** When 1 ng of ORK1 cRNA is injected into *X. laevis* oocytes a current is observed by two-electrode voltage clamp that is not present in control oocytes (Fig. 5a). In response to changes in transmembrane voltage, ORK1 currents rise to a maximum level with no apparent delay. Currents are steady with maintained voltage and show no evidence for inactivation. When external K<sup>+</sup> is in the physiologic range (i.e., 5 mM KCl solution), currents are outwardly rectifying (Fig. 5b). Under these conditions, inward currents are seen but remain small even at very hyperpolarized voltages.

ORK1 currents result from movement of K<sup>+</sup> (Fig. 5c). Oocytes expressing ORK1 and studied in solutions where KCl content is altered by isotonic substitution for NaCl exhibit reversal potentials in nearly perfect agreement with values predicted by the Nernst equation. This indicates that ORK1 currents are highly selective for K<sup>+</sup> over both Na<sup>+</sup> and Cl<sup>−</sup>. Indeed, substitution of aspartate for Cl<sup>−</sup> in the bath solution has no significant effect on currents (not shown).



**ORK1 Currents in Oocytes Are Barium Sensitive.** As predicted from growth inhibition assays (Fig. 4*a*) as well as  $^{86}\text{Rb}^+$  flux experiments (Fig. 4*c*), ORK1 currents are blocked by barium but not by TEA (Fig. 6*a* and *b*). Barium block is voltage dependent and does not change the instantaneous character of current development or lead to inactivation (Fig. 6*c*). This suggests that barium inhibits ORK1, as it does several well-characterized  $\text{K}^+$  channels (37), by direct, reversible, open-channel pore blockade. If barium does block by this mechanism, its binding energy will reflect the fraction of the electric field traversed to reach its pore site (36, 38); a determination for  $z\delta$  of  $0.80 \pm 0.02$  suggests that barium ions move through  $\approx 40\%$  of the potential drop across the membrane to bind (Fig. 6*d*).

The gene for a unique K<sup>+</sup> channel containing two P domains in tandem has been cloned from *D. melanogaster*. Cloning was achieved by complementation using a mutant strain of *S. cerevisiae* defective in K<sup>+</sup> transport (39) and a cDNA expression library produced from fly larvae (22). Channel expression in yeast cells confers uptake of K<sup>+</sup> and allows cell growth in low-K<sup>+</sup> medium; in *X. laevis* oocytes, expression yields instantaneous, K<sup>+</sup>-selective,

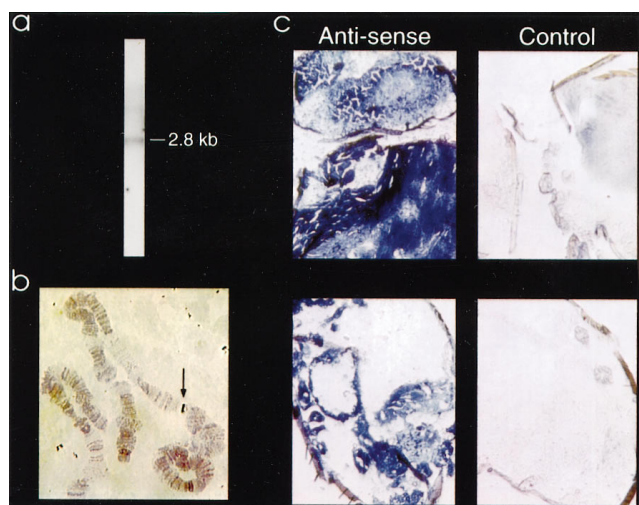


FIG. 3. ORK1 is expressed in the excitable tissues of adult *D. melanogaster*. (a) Northern blot analysis of ORK1 mRNA. *D. melanogaster* mRNA (5  $\mu$ g, adult, Clontech) was resolved in a formaldehyde/agarose gel, blotted to nitrocellulose, and probed with  $^{32}$ P-labeled 2.4-kb ORK1 *Xho*I fragment overnight at 65°C. The blot was exposed to x-ray film for 46 hr at -70°C with an intensifying screen. (b) *In situ* hybridization to polytene chromosomes identifies a band on 1-10A1-2 by the method of Todd Lavery using biotin-labeled probes produced by nick-translation and peroxidase-based visualization (Enzo Diagnostics, New York). (c) *In situ* hybridization in whole adult fly preparations reveals expression of ORK1 mRNA in excitable tissues in the head and thorax (Upper) and abdomen (Lower) with antisense but not control cRNA. Digoxigenin-labeled antisense RNA (to the terminal 300 nucleotides of the coding region and 400 nucleotides of the 3' untranslated region) or a kit-supplied control RNA were hybridized to 18- $\mu$ m sections of quick-frozen, OCT-embedded, *D. melanogaster* CS overnight at 65°C and visualized by an alkaline phosphatase-catalyzed reaction (Boehringer Mannheim).

and noninactivating currents whose rectification is altered by  $[K]_o$ . The gene and its channel have been named ORK1.

Two motifs resembling the pore-forming P domains of known  $K^+$  channels are found in ORK1 channels (Fig. 2a). In single P domain  $K^+$  channels, a "signature sequence" within the P domain (TxTxGxG, with lowercase letters indicating variable amino acid residues) is critical for  $K^+$  selectivity (2). The P<sub>1</sub> sequence of ORK1 provides a variant of this sequence (TvcsTvGyG), while the P<sub>2</sub> aligns well (TttTiGfG). As in other cloned  $K^+$  channels, each P domain is flanked by stretches of 20 or more hydrophobic amino acids (Fig. 2b, M1-M4) and ORK1 subunits are predicted to have four membrane-spanning segments (Fig. 2e). Because single P domain  $K^+$  channel subunits associate as tetramers to form functional complexes (40-42), it seems likely ORK1 channels, with two P domain subunits, will function as dimers. The hydrophilic C-terminal domain of ORK1 contains multiple consensus sites for protein kinase A and C phosphorylation and a region (residues 551-581) with strong similarity to the third intracellular loop of the somatostatin receptor subtype 3 that is thought to contribute to interaction with heterotrimeric G proteins (43). This suggests a regulatory function for this domain.

ORK1 shares its predicted two P domain, four membrane-spanning segment (2P/4TM) topology with TWIK, the product of a gene recently identified in the human expressed sequence tag data base using the BLAST alignment program (14); like ORK1, TWIK is predicted to influence membrane potential, since it yields a  $K^+$ -selective pore with extremely weak inward rectification. This 2P/4TM topology is predicted for over a dozen ORFs in the *C. elegans* genome that may be  $K^+$  channels as well (15, 16). Characterization of additional 2P/4TM channels will be required to determine if they all produce open  $K^+$ -selective channels and thus merit classification as a family based on function rather than their unique structure. While it is not yet possible to infer the

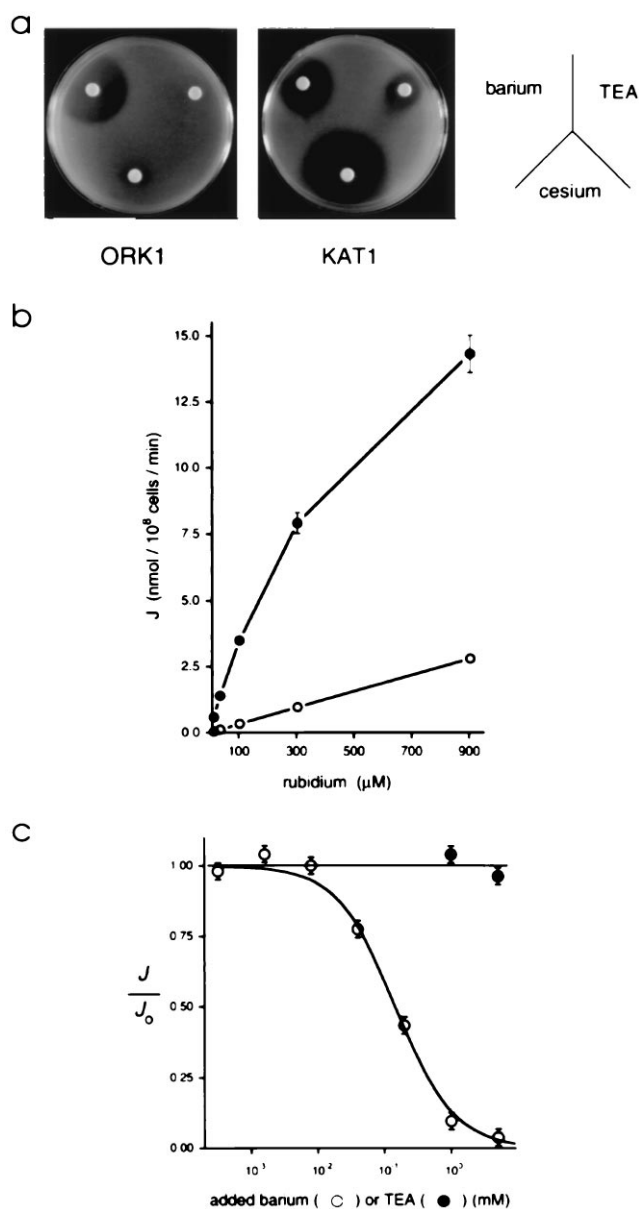


FIG. 4. ORK1-dependent yeast cell growth and  $K^+$  uptake are inhibited barium. (a) Inhibition of ORK1 or KAT1-dependent yeast growth by three common  $K^+$  channel blockers. CY162 cells ( $10^5$ ) expressing the designated channels were plated in RPD medium with 1 mM KCl. Sterile filter disks were placed on the surface of the agar and saturated with 20  $\mu$ l of solutions containing chloride salts of barium (5  $\mu$ M), TEA (20  $\mu$ M), or cesium (2  $\mu$ M). The plates were incubated for 36 hr at 30°C. (b) Uptake of rubidium by CY162 cells carrying pRAD-ORK1 (●) or pRAD (○) at various concentrations of unlabeled rubidium chloride. (c) Inhibition of rubidium uptake by barium (○) or TEA (●) at 50  $\mu$ M rubidium chloride. The solid curve is fitted to the data according to  $(1 + [blocker]/K_i)^{-1}$ , where  $K_i$  is the equilibrium dissociation constant; under these conditions 0.15 mM external barium decreases flux  $\approx 50\%$ .

evolutionary antecedents or progeny of the 2P/4TM channels, it is apparent that the P and near P segments of ORK1 are related to the human and *C. elegans* gene products as well as to Tok1 of *S. cerevisiae*, the founding member of the other two P domain  $K^+$  channel family, which has a proposed 2P/8TM topology (Fig. 2c). These alignments support the contention that a common ancestor of two P domain  $K^+$  channels was present in eukaryotes before differentiation of fungi and animals in the late Precambrian period (44). It is premature to assess the relationship between two P domain  $K^+$  channels and  $Na^+$  and  $Ca^{2+}$  channels, whose subunits carry four P domains in one contiguous polypeptide,

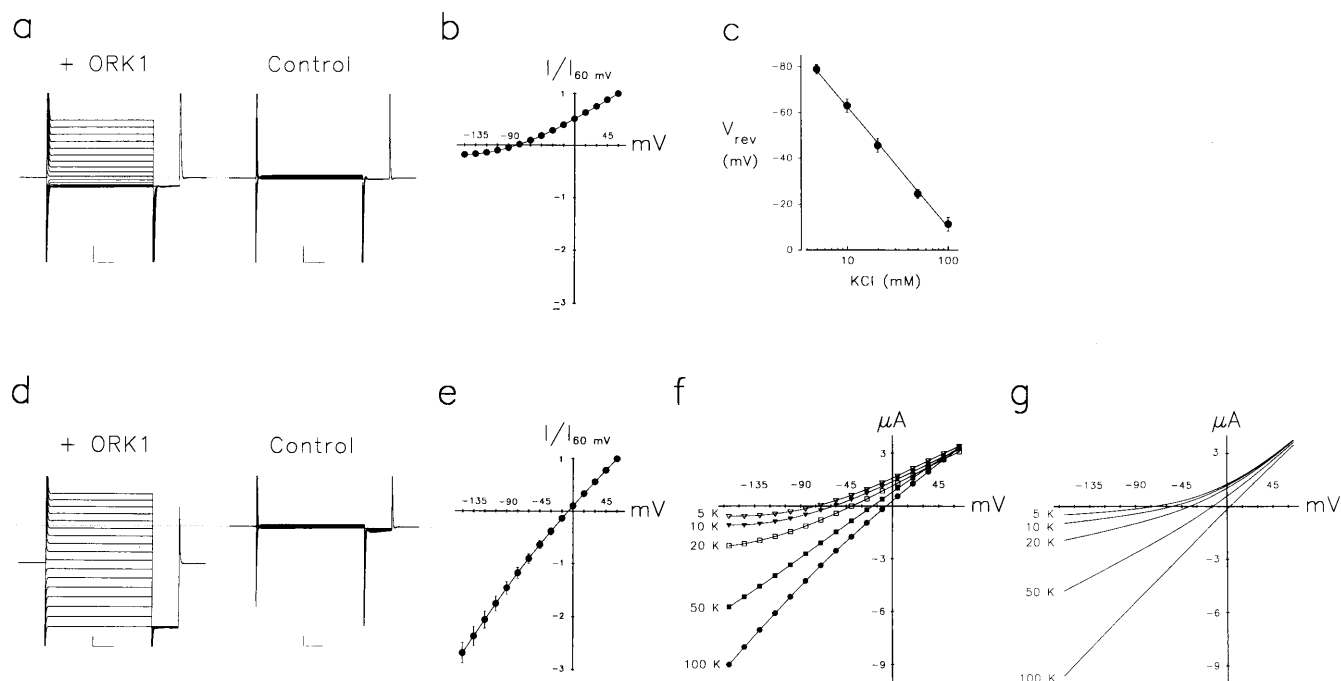


FIG. 5. ORK1 currents in *X. laevis* oocytes. (a) ORK1 currents in physiological levels of  $[K]_o$  are outwardly rectifying. Currents were assessed in oocytes injected with 1 ng of cRNA (+ ORK1) or water (control) by two-electrode voltage clamp under constant perfusion with 5 mM KCl solution. Oocytes were pulsed from  $-150$  to  $60$  mV in  $15$ -mV steps for  $75$  ms followed by a  $15$ -ms step to  $-150$  mV before returning to the holding potential of  $-80$  mV; a  $1$ -s interpulse interval was employed. Currents are displayed without leak subtraction. Scale bars represent  $2 \mu A$  and  $15$  ms. (b) ORK1 current-voltage relation in  $5$  mM KCl solution at  $10$  ms into the test pulse normalized to current at  $60$  mV by the protocol in *a* (mean  $\pm$  SEM,  $n = 4$  cells). (c) ORK1 currents are  $K^+$  selective. The reversal potential of currents was studied with  $5$ ,  $10$ ,  $20$ ,  $50$ , or  $100$  mM KCl solutions by the protocol in *a* (mean  $\pm$  SEM,  $n = 4$  cells). Linear regression gives a shift of  $55 \pm 2$  mV per  $10$ -fold change in KCl concentration. (d) ORK1 currents flow inward at hyperpolarized voltages under constant perfusion with  $100$  mM KCl solution; protocol and scale bars as in *a*. (e) ORK1 current-voltage relation in  $100$  mM KCl solution as in *b* (mean  $\pm$  SEM,  $n = 4$  cells). (f) ORK1 current-voltage relation for one oocyte studied in  $5$ ,  $10$ ,  $20$ ,  $50$ , and  $100$  mM KCl solutions as in *b*. (g) Theoretical current-voltage relations under the conditions used to study ORK1 in *e* according to Goldman (17) and Hodgkin and Katz (18):

$$I_s = P_s z_s^2 \left( \frac{VF^2}{RT} \right) \left( \frac{[S]_i - [S]_o \exp(-z_s VF/RT)}{1 - \exp(-z_s VF/RT)} \right),$$

where  $P_s$  is the permeability of  $K^+$ ,  $[S]$  refers to  $K^+$  concentration, and  $z$ ,  $V$ ,  $F$ ,  $R$ , and  $T$  have their usual meanings, and assuming an internal  $K^+$  concentration of  $90$  mM, as reported previously (35).

which are thought to have arisen through gene duplication via two P domain channel intermediates (45).

The effects of extracellular  $K^+$  on ORK1 appear to result directly from unequal permeant ion concentrations across the membrane, as expected from constant field theory for an open  $K^+$  channel (17, 18). This type of instantaneous rectification through open  $K^+$  (and  $Na^+$ ) channels has been recognized in nerves and muscles previously (20, 46, 47) and differs from other known effects of  $[K]_o$  on  $K^+$  channel gating (48–53) and blockade (54–57). The effects of  $[K]_o$  on ORK1 are also distinct from those observed with Tok1, the archetype of the other two P domain  $K^+$  channel family. Tok1 allows only small inward currents in intact cells, even when extracellular  $K^+$  is high, but it does alter the voltage at which it passes outward currents in response to changes in  $[K]_o$  (13). In contrast, ORK1 passes large inward  $K^+$  currents in elevated  $[K]_o$ . Tok1 is thus categorized as an “outward rectifier” (13), in analogy to the inward rectifier  $K^+$  channels that shift the voltage at which they pass inward currents in response to changes in  $[K]_o$ , and ORK1 is classified as an “open rectifier.” Of note,  $K^+$  conductances that are sensitive to  $[K]_o$  have a recognized role in normal physiology (58) as well as the pathogenesis of epilepsy and postischemic cardiac arrhythmias (59, 60) but the channels that mediate these effects are undefined.

The attributes of ORK1 currents ( $K^+$  selectivity,  $[K]_o$ -sensitive rectification, voltage independence, and TEA insensitivity), are similar to those identified with the “residual leak” conductance hypothesized to mediate the resting potential of myelinated nerves dissected from vertebrates and, thus, indirectly argues for

the feasibility of that model (21). Study of ORK1 in flies appears to offer a powerful strategy to explore the role of “leak” conductances in nervous function (61). ORK1 might be expected to have two effects in the excitable tissues of *D. melanogaster*. First, ORK1 yields a  $K^+$ -selective current across all voltages studied which will shift resting membrane potential toward the equilibrium reversal for  $K^+$  ( $E_K$ ). Second, rectification of ORK1 currents is  $[K]_o$  dependent, and this will influence the magnitude and duration of action potential afterhyperpolarizations and firing rates. Since action potential repolarization results from outward flux of  $K^+$  leading to transient elevation in  $[K]_o$  at the external face of the membrane (62, 63), it follows that the size of inward currents through ORK1 channels will exhibit  $[K]_o$ -dependent hysteresis with each wave of excitation and, so, dynamically influence the kinetics of changes in membrane potential. These effects would affect (and may control) overall membrane excitability. The potential of a genetic approach in flies is underscored by observation that mutations in voltage-gated ion channels have proven a rich source of *D. melanogaster* behavioral mutants (4–6, 64).

Successful application of the complementation cloning method that isolated ORK1 has previously been restricted to channels and transporters from plants (23, 65). In general, plants and fungi maintain resting membrane potentials more negative than  $-100$  mV. Under these conditions, most voltage-gated  $K^+$  channels from animals are closed. Functional expression of ORK1 in yeast cells may result, first, from its ability to pass inward currents at negative potentials and, second, from its similarity to Tok1, an



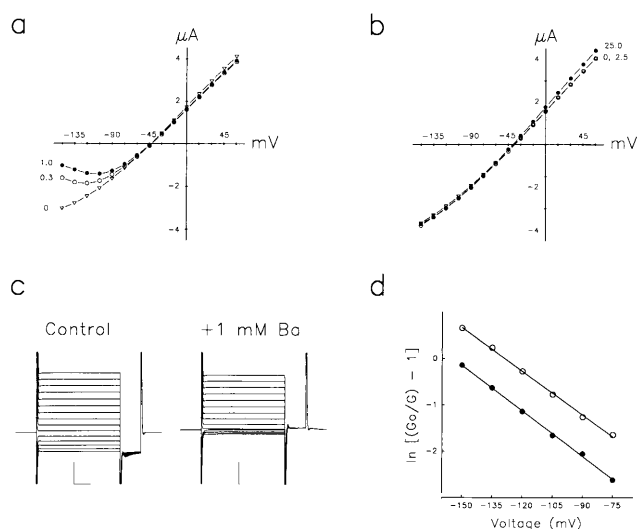


FIG. 6. ORK1 currents in oocytes are blocked by barium but not TEA. (a) Current-voltage relationship with constant perfusion of 20 mM KCl solution containing 0 mM (▽), 0.3 mM (○), or 1 mM (●) barium chloride by the protocol in Fig. 4a (mean  $\pm$  SEM,  $n = 3$  cells). (b) Current-voltage relationship with constant perfusion of 20 mM KCl solution with 0 mM (▽), 2.5 mM (○), or 25 mM (●) TEA chloride isotonicity substituted for NaCl (mean  $\pm$  SEM,  $n = 3$  cells). (c) ORK1-induced currents in an oocyte perfused with 20 mM KCl solution in the absence (control) or presence of 1 mM barium chloride by the protocol in Fig. 4a. Scale bars represent 2  $\mu$ A and 15 ms. (d) Voltage dependence of barium block was assessed by measurements of conductance from  $-150$  mV to  $-75$  mV in the presence of 0.3 mM (○) or 1 mM (●) barium chloride by the protocol in Fig. 4a. The data shown are from one oocyte but are representative.  $G_0$  and  $G$  are the cord conductances measured in the absence and presence of barium, respectively. The electrical distance was calculated from  $K_i(V) = K_i(0)\exp(z\delta FV/RT)$ , where  $K_i(0)$  is the zero voltage inhibition constant,  $z$  is the valence of the blocking ion, and  $\delta$  is the fraction of the applied voltage drop experienced at the binding site (36).

endogenous yeast two P domain channel that is processed and translocated to the cell membrane (66). Expression of ORK1 in yeast makes possible genetic approaches to ion channel structure and function not possible in other expression modalities (25, 67).

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